

STRAIN DIFFERENCES IN THE EFFECT OF LONG-TERM TREATMENT WITH MELATONIN ON KAINIC ACID-INDUCED STATUS EPILEPTICUS, OXIDATIVE STRESS AND THE EXPRESSION OF HEAT SHOCK PROTEINS

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ABSTRACT

The present study compared the effects of subchronic treatment with melatonin administered via subcutaneous osmotic minipumps for 14 days (10 mg/kg per day) on kainic acid (KA)-induced status epilepticus, oxidative stress and expression of heat shock protein (HSP) 70 in the frontal cortex and hippocampus between normotensive Wistar rats and spontaneously hypertensive rats (SHRs). SHRs showed increased lipid peroxidation (LP) in the cortex and hippocampus and decreased cytosolic superoxide dismutase (SOD/CuZn) production in the cortex compared to Wistar rats. Long-lasting seizures induced by KA (12 mg/kg, i.p.) were accompanied by increased LP and expression of HSP 70 in the hippocampus of the two strains and increased SOD/CuZn production in the frontal cortex of SHRs. Pretreatment with melatonin failed to suppress the KA-induced SE in the two strains though the latency for seizure onset was significantly increased in SHRs. The increased LP induced by KA in the hippocampus was attenuated by melatonin pretreatment both in Wistar rats and SHRs. The increase of SOD/CuZn and mitochondrial SOD/Mn production was strain- and area-specific in melatonin- KA treated groups. Melatonin prevented the KA-induced increased expression of HSP 70 in the hippocampus of KA-treated Wistar rats. The study suggests that the potential efficacy of melatonin pretreatment on SE-induced oxidative stress and neurotoxicity is strain- and area-specific.

Key words: *Kainic acid; melatonin; oxidative stress; heat shock protein; spontaneously hypertensive rats; Wistar rats.*

Introduction

Epilepsy has been described as a condition of excessive neuronal discharge associated with or resulting from oxidative stress (Shin et al., 2011; Waldbaum and Patel, 2010). Epileptic biomarkers, such as catalytical antioxidants and oxidative products in neuronal tissues have been monitored for evaluation of the degree of epileptic pathogenesis. The neurotoxin kainic acid (KA) triggers neuropathologic cellular changes in the hippocampus characterized by an overloading of intracellular calcium, mitochondrial membrane rupturing, activation of intracellular enzyme cascades, including the nitric oxide synthase and increased levels of free radicals/reactive oxygen species (ROS) from the mitochondrial intermembrane space into the cytosol (Srivastava et al., 2008). The formation of free radicals results in an extensive lipid peroxidation, which damage cellular organelles and membranes, and finally leads to cell death. Thus, oxidative stress resulting from excitotoxicity is suggested to play a critical role in epileptic brain damage (Bondy and Lee, 1993). Harmful changes in cells, including calcium influx and generation of ROS induce or suppress the expression of genes in neurons thereby influencing synthesis of proteins (Rajdev and Sharp, 2000). The expression of stress proteins, referred to members of heat shock proteins (HSP) is caused by a variety of injurious stimuli in the brain and is also considered as an appropriate marker of excitotoxicity. Several studies suggested that there exist a close relationship between ROS and the expression of members of the Hsp70 family (Ambrosio et al., 1995; Kukreja et al., 1994; Lee J. Y.

and Corry, 1998). In addition, the increased expression of HSP 70 was seen after KA-induced SE in rat brain (Gupta and Briyal, 2006).

The disturbance in the levels of the antioxidant enzymes is a crucial step involved in dysregulation of physiological processes implicated in the pathogenesis of arterial hypertension (Harris, 1992). Experimental data demonstrate that the direction of changes in the activity of antioxidant system strongly depends on the severity of oxidative stress. In this regard, moderate levels of ROS production are able to enhance the activity of the defense system by stimulation of different antioxidant enzymes (Vogt, 1998). However, when the oxidative load exceeds the defense potential, the adaptive response of the antioxidant system could be disturbed (Csonka et al., 2000). It has been reported that the expression of antioxidant enzymes is increased in the myocardium of spontaneously hypertensive rats (SHRs) following the induction of oxidative stress (Csonka et al., 2000). However, the reduction of the defense antioxidant enzymatic activities in SHRs compared to normotensive Wistar Kyoto rats indicates a disturbance of the defence system as a sequence of the enhanced oxidative stress in the SHR model (Polizio and Peña, 2005). Currently, number of protective approaches with potential antioxidants has been applied to improve hypertension (Levonen et al., 2008; Schiffrin et al., 2010). In addition to a widely accepted experimental model of essential hypertension, SHRs could be considered as a tool for studying the link between hypertension and epilepsy (Greenwood et al., 1989; Scorza et al., 2005; Tchekalarova, 2010, 2011). Furthermore, substantial data support the view point that some of the physiological and biochemical markers of epilepsy are also evident in naive SHRs. Thus, hippocampal neuropathology, including neuronal loss, mostly in the CA₁ subfield, and astrocyte reactivity has been reported in intact adult SHRs (Pietranera et al., 2006; Sabbatini et al., 2000). Vorobyov and co-authors (2011) found that the EEG spectral profiles are similar in SHRs suffering from congenital hypertension and in KA-treated normotensive rats. In this regard, we have shown that naive and epileptic SHRs exhibit similar anxiety level associated with low level of serotonin and dopamine in the hippocampus (Tchekalarova et al., 2011). Moreover, the naïve SHRs were characterized with abnormal behavioral responses and biochemical parameters, which are also characteristic for epileptic rats (Tchekalarova et al., 2011).

Clinical evidence has revealed that melatonin is implicated in epilepsy, it is able to reduce the spiking activity and seizure frequency in patients with intractable epilepsy (Anton-Tay, 1974; Molina-Carballo et al., 1997) and it is detected in high levels during the postictal period (Bazil et al., 2000). Melatonin possesses a low toxicity and may be used for seizure control in conjunction with anti-seizure medications (Rufo-Campos et al., 2002). Melatonin has been also characterized as a potent anticonvulsant in a number of seizure models in rodents, including acute seizure tests (Albertson et al., 1981; Borowicz et al., 1999; Lapin et al., 1998; Yamamoto and Tang, 1996) and models of epilepsy (Albertson et al., 1981; Mevissen and Ebert, 1998; Tchekalarova et al., 2013). It has been shown to exert neuroprotective effects against neuronal damage in animal models of neurotoxicity i.e. stroke and traumatic brain injury (Chung, 2003) as well as toxic quinones and oxidative stress produced by catecholamines (Hirata et al., 1998). Single injection of melatonin in rats before and during the KA- or pilocarpine-induced status epilepticus (SE) has neuroprotective effect by reducing the neuronal death, supragranular mossy fiber sprouting, lipid peroxidation (LP), and microglial activation (Banach et al., 2011; de Lima et al., 2005; Guisti et al., 1996). So far, there has been accumulated a broad spectrum of *in vitro* and *in vivo* studies confirming the attitude that melatonin behaves as a free radical scavenger and potent antioxidant (reviewed in: Russel et al., 2000). Although the efficacy of a single injection of melatonin against oxidative stress and KA-induced seizures has been already studied in mice (Mohanan and Yamamoto, 2002), there is none study focused on comparative investigation of the putative role of long-term melatonin exposure against the KA-induced excitotoxicity during the acute period between SHRs and normotensive Wistar rats.

The brain is particularly sensitive to attacks of ROS. The frontal cortex and the hippocampus are connected with each other through different neurotransmitter systems and have been proposed to be particularly vulnerable to KA-induced SE (Ben-Ari et al., 1980; Schwob et al., 1980; Sperk et al., 1983; Chen and Buckmaster, 2005). We hypothesized that the rate of oxidative damage during SE may vary in a region- and melatonin- specific manner in Wistar rats and SHRs. Therefore, the aim of the present investigation was to check out and compare the efficacy of subchronic melatonin exposure on KA-induced seizure severity and changes in the LP, enzymatic antioxidant defense systems and heat shock protein (HSP) 70 expression in the frontal cortex and hippocampus at 4 hour (h) following SE in normotensive Wistar rats and SHRs.

2. Materials and methods

2.1. Animals

The experiments were performed on adult male normotensive Wistar rats obtained from the animal facility of the Bulgarian Academy of Sciences and spontaneously hypertensive rats (SHRs) from the local breeding house (Medical University, Sofia). The rats weighing 180-200 g were adapted for one week under standardized laboratory conditions (12 h/12 h light/dark cycle, temperature $22\pm 2^{\circ}\text{C}$, 50 % relative humidity) in groups of 2 in plastic cages with soft bedding. Food and water were available ad libitum throughout the study except during the tests. The experimental protocol was in compliance with the European Communities Council Directive of 24th November 1986 (86/609/EEC) and the experimental design was approved by the Institutional Ethics Committees of Sofia Medical University and the Institute of Neurobiology for the National Science Fund grant DTK 02/56 2009-1012.

2.2. Experimental design

The animals were divided into eight experimental groups (n=10) as follows: Group I: Naive and sham normotensive Wistar group treated with vehicle (Wis-C-veh); Group II: control+melatonin Wistar group (Wis-C-mel); Group III: Kainic acid (KA) group (Wis-KA-veh); Group IV: KA + melatonin group (Wis-KA-mel); Group V: Naive and sham spontaneously hypertensive group (SHRs-C-veh) treated with vehicle; Group VI: control+melatonin SHR group (SHRs-C-mel); Group VII: KA group (SHRs-KA-veh); Group VIII: KA + melatonin group (SHRs-KA-mel). Melatonin (Sigma-Aldrich, Bulgaria) was applied chronically for a period of two weeks via osmotic minipumps at a dose of 10 mg/kg/day. Alzet osmotic minipumps were filled with drug dissolved or vehicle (0.9% NaCl and DMSO (2:1), pumping rate 0.5 $\mu\text{l/h}$, information provided by manufacture). There are not established any differences between naive and sham rats. A method of melatonin infusion via s.c. osmotic minipumps provided constant steady-state hormonal concentrations.

2.3. Arterial blood pressure measurements

Systolic arterial blood pressure (ABP) was measured non-invasively in conscious unrestrained SHRs by a tail cuff method (Ugo Basile Blood Pressure Recorder 5800) before the start of experimental procedures. The ABP value for each rat was calculated as mean of three measurements.

2.4. Kainic acid-induced status epilepticus

On the 14th day of vehicle/melatonin s.c. infusion animals from groups III, IV, VII and VIII received intraperitoneal injection of KA (Ascent Scientific, UK) at a dose of 12 mg/kg dissolved in sterile saline (0.9 % NaCl) or saline (groups I, II, V, VI) in a volume of 1 ml/kg of body weight. The protocol used to elicit KA-induced SE was based on previous studies (López-Meraz et al., 2005; Morales-Garcia et al., 2009). After injection of KA, the animals were put in individual cages and observed for 4 h to evaluate the appearance of seizures. The intensity of seizures was assessed

according to the Racine's scale (1972) consisting of six stages (0-5), which correspond to the successive developmental stages of motor seizures: (0) normal non-epileptic activity; (1) facial automatism, sniffing, scratching, wet dog shakes; (2) head nodding, staring, tremor; (3) forelimb clonus with lordotic posture; (4) rearing and continued forelimb clonus, salivation; (5) forelimb clonus and loss of posture. Latency for the onset of the first forelimb clonus with lordotic posture was also evaluated.

2.5. Biochemical experiments

Biochemical tests were conducted 4 h after KA injection. The animals were sacrificed by decapitation under a light anesthesia (CO₂). Brains were quickly dissected on ice and the frontal cortex and the hippocampi were bilaterally removed. The tissue samples were frozen in liquid nitrogen, and stored at -70°C before analysis.

2.4.1. Measurement of lipid peroxidation

The extent of lipid peroxidation was determined quantitatively by direct measurement of hydroperoxides in redox reactions with ferrous ions. Therefore the tissue samples were homogenized in cold 20 mM HEPES buffer (pH 7.2) and extracted with chloroform. The extracted lipid peroxides were assayed with LPO assay kit (Cayman Chemical Company, USA) according the instructions provided. The resulting ferric ions were detected using thiocyanate ion as the chromogen and by reading the absorbance at 500 nm. The extent of lipid peroxidation was expressed as nmol.

2.4.1. Measurement of cytosolic and mitochondrial superoxide dismutase

The tissue samples were homogenized in cold 20 mM HEPES buffer (pH 7.2) and centrifugated at 1500 × g for 5 min, at 4 °C. To separate cytosolic and mitochondrial SOD, the 1500 × g supernatant was again centrifuged at 10 000 × g for 15 min, at 4 °C. The resulting supernatant was tested for cytosolic SOD and the pellet – for mitochondrial SOD

with SOD assay kit (Cayman Chemical Company, USA) The results were expressed as U/ml.

2.4.2. Heat shock protein

Western Blotting

Tissues were washed once in ice-cold PBS and homogenized in 5 ml of cold 20 mM HEPES buffer, pH 7.2, containing 1 mM EGTA, 210 mM mannitol and 70 mM sucrose. Protein concentration was determined by spectrophotometric measuring of the homogenates at 280 nm. Equal amounts (20 mg/lane) of protein samples were run on an 12 % SDS polyacrylamide gel. The proteins were transferred onto nitrocellulose membrane and blocked with 3% bovine serum albumin in TBS-0.05% Tween. The membrane was incubated with the primary mouse anti-Hsp72 antibody μ chain (Invitrogen) 1:500, for 2 h at room temperature or overnight at 4°C. The membrane was washed 3 times with TBS-Tween and further incubated in the secondary antibody anti-mouse μ chain, raised in goat and conjugated with alkaline phosphatase (Vector Labs, USA) 1: 250. After 3 times washing in TBS-Tween the membrane was incubated in 10 ml of ABC-AmP reagent (Vector Labs, USA) for 10 min at room temperature and washed again. Then the membrane was equilibrated in TBS for substrate (pH 9.5) and incubated in substrate solution BCIP/NBT (Vector Labs, USA) at room temperature for about 30 min. After developing of appropriate density color bands the membrane was rinsed in PBS and air dried. On every SDS-PAGE one lane was extrapolated to the same standard of 20 mg of control rat brain tissue protein and all other bands on each gel were expressed relative to this standard. Blots were scanned and analyzed with the ImageJ software (V 1.42q).

2.5. Statistical analysis

The data are expressed as mean±S.E.M. The seizure severity scores following KA injection were evaluated by means of two-way ANOVA with subsequent comparison with Kruskal-Wallis test while the biochemical parameters by means of three-way ANOVA (SigmaStat® SPSS). The incidence of seizures and mortality was evaluated by Fisher's exact test. The level of statistical significance was set at 5 %.

3. Results

Control SHR's showed significantly higher ABP (181±1.45 mm Hg, $p<0.005$) in comparison with the normotensive Wistar controls (134±1.45 mm Hg).

3.1. Effect of subchronic melatonin pretreatment on KA-induced seizures

As shown in Table 1, there were no behavioral differences between Wistar rats and SHR's pretreated with either vehicle or melatonin (10 mg/kg for 14 days via s.c. osmotic minipumps). The systemic i.p. injection of a single excitotoxic dose of KA (12 mg/kg) led to development of progressive motor changes and SE both in Wistar rats and SHR's similarly to previously described (Tchekalarova et al., 2010). Most of the KA-treated animals were characterized with facial automatisms, wet dog shakes and head nodding (partial seizures) during the first hour of observation. During the second and the third hour following KA administration, this activity progressed to secondary generalized seizures i.e. forelimb clonus with lordotic posture followed by rearing and forelimb clonus and loss of posture. The Racine's score reached 3.8±0.32 points in Wistar rats and 3.7±0.47 points in SHR's, respectively. The latency for onset of the first clonic seizure induced by KA injection was significantly lower in SHR's-KA-veh group compared to Wistar-KA-veh group ($^{\circ}p=0.02$) (Table 1). Subchronic melatonin pretreatment significantly increased the latency for the onset of the first clonic seizure in SHR's ($*P=0.034$) (Table 1).

3.2. Effects of melatonin treatment on the level of lipid peroxidation

Lipid peroxidation as a marker of oxidative stress showed strain-dependent differences both in frontal cortex ($\#p<0.001$) and hippocampus ($\#p<0.001$). SHR's have higher level of lipid peroxidation in naïve and melatonin treated controls ($\#p<0.001$), as well in KA- vehicle ($\#p<0.001$) and KA- melatonin groups ($\#p=0.066$) compared to Wistar rats in frontal cortex. KA-treatment provoked a significant increase in the hippocampal lipid peroxidation in Wistar rats ($*p=0.028$), which level was dramatically decreased even below the control level after sub-chronic melatonin treatment ($^{\circ}p<0.001$). KA-treated SHR's also displayed an increased oxidative stress in the hippocampi ($*p=0.008$), which was abolished by melatonin pretreatment ($^{\circ}p<0.001$) (Fig.1). Similar drug-induced changes were not found in the frontal cortex.

3.3. Effects of melatonin on the cytosolic superoxide dismutase (SOD Cu/Zn) activity in the frontal cortex and the hippocampus of Wistar and SHR's

Naïve SHR's showed lower SOD Cu/Zn level in the frontal cortex compared to respectively Wistar controls ($\#p<0.001$). KA-treatment increased enzyme level only in SHR's frontal cortex ($*p<0.001$). Although melatonin pretreatment abolished KA-induced SOD Cu/Zn increase in SHR's frontal cortex ($^{\circ}p<0.001$), on the other hand the hormone treatment raised SOD Cu/Zn cortical level in KA-treated Wistar ($^{\circ}p<0.001$), and hippocampal ($^{\circ}p<0.001$) enzyme level in both strains as well as in naïve SHR's ($*p=0.016$) (Fig 2).

3.4. Effects of melatonin on the mitochondrial superoxide dismutase (SOD Mn) activity in the frontal cortex and the hippocampus of Wistar and SHR's

Neither strain differences nor KA-induced changes in SOD Mn were found in controls. Melatonin treatment, however showed dual effect – it decreased cortical SOD Mn level both in

naïve (#p=0.024) and KA-treated SHR's (#p<0.001), but increased the enzyme level in the hippocampi of KA-treated Wistar rats ($^{\circ}$ p= 0.020) (Fig. 3).

3.5. Effects of melatonin on the heat shock protein 70 in the frontal cortex and the hippocampus of Wis and SHR's

KA-treatment increased HSP70 level in hippocampus of Wistar rats (*p<0.001) and in SHR's hippocampus (*p= 0.003), but this effect was weaker in SHR's (#p=0.001). SHR's frontal cortex remains unaffected by the neurotoxin. Melatonin treatment was able to abolish only KA-induced increase in HSP70 level in the hippocampi of Wistar rats ($^{\circ}$ p=0.05) (Fig. 4).

Discussion

In the present study, the subchronic melatonin treatment increased the latency of KA-induced seizures in SHR's but not in normotensive Wistar rats. Similarly, administration of melatonin at a dose of 10 mg/kg for 60 days, starting three hours after the beginning of SE, exerted a more efficient attenuation of spontaneous recurrent seizures in SHR's than in Wistar rats because it was able to suppress the seizure activity after discontinuation of the melatonin treatment in SHR's (Tchekalarova et al., 2013; Petkova et al., submitted). We can suggest that the higher efficacy of melatonin treatment on seizure activity in SHR's than in Wistar rats might be related to simultaneous decrease of the blood pressure detected in epileptic SHR's (Petkova et al., unpublished). There are emerging experimental and clinical studies considering the close relationship between hypertension and epilepsy (Tomson et al., 1998; Hilz et al., 2002; Devinsky et al., 2004). However, melatonin failed to suppress the development of KA-induced SE both in Wistar rats and SHR's. These results are in accordance with our previous works demonstrating that the long-term melatonin treatment after KA-induced SE decreased the latency for onset of the first spontaneous recurrent seizure and attenuated the seizure frequency during the treatment period without preventing the development of chronic epileptic state in Wistar rats and SHR's (Thekalarova et al., 2013; Petkova et al., unpublished). Although the majority of data indicate anticonvulsant activity of melatonin in both animal models and in patients with epilepsy, this drug was only suggested for add-on therapy in epileptic patients with insomnia. The contradictory literature data concerning the anticonvulsant efficacy of melatonin applied at pharmacological doses are related to its time-, age- and model-dependent effect. Thus, although a single injection of melatonin exerts an anticonvulsant activity in different seizure tests (Banach et al., 2011; De Lima et al., 2005), experimental data suggest that the time of administration, the duration of treatment and the age of the testing subjects are very important for drug efficacy (Costa-Lotufo et al., 2002; Musshoff and Speckmann, 2003). Costa-Lotufo and co-authors (2002) reported that subchronic (10-50 mg/kg, i.p. for one week in the morning and in the afternoon, respectively) but not acute melatonin treatment prevented the pilocarpine-induced seizure activity. Furthermore, acute melatonin injection (20 mg/kg, ip) was reported to be ineffective to PTZ and KA-induced seizures in rats (Xu and Stringer, 2008) whereas the same design of melatonin treatment suppressed the KA-induced seizures in mice and mitochondrial DNA damage in the mouse brain cortex (Mohan and Yamamoto, 2002). Our treatment protocol provided constant steady-state drug concentration and the time of treatment could be neglected. Alternatively, our results suggest that the anticonvulsant efficacy of melatonin depends on the seizure model and co-morbid hypertension.

In accordance with several other studies (Dal-Pizzol, 2000; Gupta et al., 2002; Marini et al., 2004), we demonstrated that KA-induced SE is accompanied by increased LP level in hippocampus homogenate of SHR's and Wistar rats after 4 hours of acute phase of seizures. Our data confirm the suggestion that ROS are involved in the mechanism of neurotoxicity triggered by KA during the acute phase in SHR's and normotensive Wistar rats. Moreover, literature data demonstrated that the increased brain LP could be detected also during the late periods (24-72 h after SE) (Candelario-Jalil et al., 2001; Dal-Pizzol, 2000; Kubera et al., 2004). Our results are in support of previous

finding that single melatonin injection is able to prevent the KA-induced LP augmentation in mice (Mohan and Yamamoto, 2002). Although a large body of *in vivo* and *in vitro* evidence revealed that melatonin function as a free-radical scavenging antioxidant (reviewed in: Galano et al., 2011), in our study melatonin failed to decrease the LP level below the basal value in naive SHR and normotensive Wistar rats. Indeed, Gönenç et al. (2005) revealed that melatonin administration alone decreased the level of LP in the hippocampus but not in the frontal cortex compared to control rats. These discrepancies could result from variations in experimental species, different melatonin doses and routes of administration.

In our study, the markers of oxidative stress, which showed an increased LP and decreased SOD Cu/Zn activity in a model of essential hypertension compared to Wistar rats indicates an enhanced oxidative stress in SHR. These results confirm previous findings that naive SHR are characterized by a disturbed oxidative defence system compared to Wistar Kyoto rats in physiological conditions (Polizio and Peña, 2005). The reported divergence of the defence system as a sequence of enhanced oxidative stress in the SHR is in favor of the assumption that the levels of antioxidant enzymes are crucial for dysregulation of physiological processes implicated in the pathogenesis of arterial hypertension (Harris, 1992). SOD is one the most important antioxidant enzyme, which protects against oxidative damage by catalyzing the dismutation of superoxide anion to hydrogen peroxide, thus contributing to decreased formation of hydroxyl radical formation (Coyle and Puttfarcken, 1993). In vertebrates, copper/zinc-containing SOD (SOD Cu/Zn) and manganese-containing (SOD Mn) are the predominant isoforms found either in the cytosol or the mitochondrial matrix, respectively. Mitochondrial dysfunction and ROS localized there plays a crucial role in the mechanisms leading to neuronal cell death during epileptogenesis (Kunz, 2002). Several studies demonstrated KA-induced disturbance in the mitochondrial function in rodents (Liang and Patel, 2006; Milatovic et al., 2001). We found that KA provoked an enhancement of the cytosolic SOD Cu/Zn 4 hours after SE only in the frontal cortex of SHR. Literature data showed that seizures and SE could alter oxidative stress by either activation or suppression of free radicals scavenging enzymes such as SOD in different brain areas (reviewed in: Devi et al., 2008). The increased cytosolic SOD activity in SHR might reflect the higher seizure susceptibility following KA injection of SHR compared to Wistar rats. Literature data support the presumption that seizures provoked changes in the oxidative defence system are influenced by the previous level of oxidative stress, brain area, strain used and time points detected for the direction of changes. Thus, Candelario-Jalil (2001) revealed that systemic administration of an excitotoxic dose of KA decreased the hippocampal SOD activity with respect to basal levels detected 24 h after KA application. On the other hand, the increased hippocampal LP after KA or pilocarpine-induced SE in female Wistar rats were more pronounced at 12-14 h after SE but returned to basal level in KA model or decreased in pilocarpine model 7-9 days or 75-80 days after the end of SE (Dal-Pizzol et al., 2000). Antioxidant-like melatonin increased cytosolic and mitochondrial SOD activity in the frontal cortex and the hippocampus but the efficacy of this hormone was higher in normotensive Wistar rats than SHR. The observed decrease in the LP level and increased SOD activity after melatonin pretreatment in KA-induced SE confirm the broadly accepted assumption that melatonin function as a free-radical scavenging antioxidant and inhibitor of LP (reviewed in: Yonei et al., 2010). However, comparison of the anticonvulsant and antioxidant efficacy of melatonin in the two strains suggests lack of a direct link between the anticonvulsant and antioxidant efficacy of melatonin.

A tendency for increased expression in the frontal cortex and a significant upregulation in the hippocampus of HSP 70 was detected both in Wistar rats and SHR after KA-induced SE. These results agree with previous findings that SE produced by systemic KA induced an expression of HSP 70 in neurons known to be susceptible to this neurotoxin and in the hippocampus, in particular (Gonzalez et al., 1989; Vass et al., 1989). Furthermore, a strong correlation between the duration of SE and the degree of HSP 70 expression was suggested by some authors (Lowenstein et al., 1990;

Shimosaka et al., 1992). Recently, several reports demonstrated increased sensitivity to heat-induced HSP 70 in vascular smooth muscle cultures, vibroblast cultures and other tissue in genetically hypertensive rodents (Hamet et al., 1990 A,B; Lukashev et al., 1991). In addition, an increased transcription of many HSP family genes, including HSP60, HSP70 and HSP90, and heat shock factor-1 have been reported in hypertensive rats (Zhou et al., 2005). In the present study we established that melatonin pretreatment over 14 days via s.c. implanted osmotic minipump prevented the KA-induced increased expression of HSP 70 in the hippocampus of Wistar rats. Curiously, melatonin failed to prevent the increased expression of HSP 70 in a model of essential hypertension. We can suggest that the low efficacy of melatonin is related to the fact that in SHR the cellular homeostasis and defence antioxidant enzyme system is disturbed in naive SHR compared to normotensive Wistar rats. The observed decreased LP level as well as the enhanced cytosolic and mitochondrial SOD, which are accompanied by a decreased expression of the HSP 70 in the hippocampus of Wistar rats pretreated with melatonin support the suggestion that there exist a close relationship between ROS and the expression of members of the Hsp70 family (Kukreja et al., 1994; Ambrosio et al., 1995; Lee J. Y. and Corry, 1998).

The concentration of melatonin is higher in brain ventricles than in the peripheral plasma following its exogenous administration (Tan et al. 2010). Because of its proximity to the ventricles, the hippocampus is one of the most peculiar brain structures, which may be susceptible to the action of melatonin (El Sherif et al. 2002). This may explain why in our study melatonin was more effective in the hippocampus than the prefrontal cortex as concern the changes in LP, cytosolic and mitochondrial SOD and HSP 70. According to these findings, the pattern of oxidative injury induced by KA seems to be highly region-specific.

In conclusion, SHR showed increased seizure susceptibility and disturbed defense antioxidant system compared to normotensive Wistar rats. Subchronic systemic melatonin treatment exerted a mild anticonvulsant effect following KA injection in SHR but not in Wistar rats. However, the efficacy of melatonin in preventing the KA-induced changes in the markers of oxidative stress and neurotoxicity was more pronounced in Wistar rats than in SHR suggesting a lack of a direct link between the seizure activity and these markers.

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REFERENCES

1. Albertson TE, Peterson SL, Stark LG Lakin ML Winters WD, The anticonvulsant properties of melatonin on kindled seizures in rats. *Neuropharmacol* 1981; 20: 61– 66.
2. Ambrosio G, Tritto I, Chiariello M. The role of oxygen free radicals in preconditioning. *J Mol Cell Cardiol.* 1995 Apr;27(4):1035-9.
3. Anton-Tay F (1974), Melatonin: effects on brain function. *Adv Biochem Psychopharmacol* 11:315– 324.
4. Banach M, Gurdziel E, Jêdrych M, Borowicz K, Melatonin in experimental seizures and epilepsy. *Pharmacol Rep* 2011; 63:1-11.
5. Bazil CW, Short D, Crispin D, Zheng, W (2000), Patients with intractable epilepsy have low melatonin, which increases following seizures. *Neurology* 55:1746–1748.
6. Ben-Ari Y, Tremblay E, Ottersen OP, Meldrum BS. The role of epileptic activity in hippocampal and "remote" cerebral lesions induced by kainic acid. *Brain Res.* 1980 Jun 2;191(1):79-97.
7. Borowicz KK, Kaminski R, Gasior M, Kleinrok Z, Czuczwar SJ, Influence of melatonin upon the protective action of conventional anti-epileptic drugs against maximal electroshock in mice. *Eur Neuropharmacol* 1999; 9:185–190.

8. Bondy SC, Lee DK. Oxidative stress induced by glutamate receptor agonists. *Brain Res.* 1993 610:229-33.
9. Candelario-Jalil E, Al-Dalain SM, Castillo R, Martínez G, Fernández OS. Selective vulnerability to kainate-induced oxidative damage in different rat brain regions. *J Appl Toxicol.* 2001, 21: 403-407.
10. Chen S, Buckmaster PS. Stereological analysis of forebrain regions in kainate-treated epileptic rats. *Brain Res.* 2005 1057:141-52.
11. Chung SY, Han SH, Melatonin attenuates kainic acid-induced hippocampal neurodegeneration and oxidative stress through microglial inhibition. *J Pineal Res* 2003; 34: 95-102.
12. Costa-Lotufo LV, Fonteles MM, Lima IS, de Oliveira AA, Nascimento VS, de Bruin VM, Viana GS, Attenuating effects of melatonin on pilocarpine-induced seizures in rats. *Comp Biochem Physiol C Toxicol Pharmacol* 2002; 131: 521-529.
13. Coyle JT, Puttfarcken P. Science. 1993 Oxidative stress, glutamate, and neurodegenerative disorders. *Science.* 262: 689-695.
14. Csonka C., Pataki T., Kovacs P., Muller SL., Schroeter ML., Tosaki A., Effects of oxidative stress on the expression of antioxidative defense enzymes in spontaneously hypertensive rat hearts. *Free Rad. Bio. Med.* 2000, 29(7), 612-619
15. Dal-Pizzol F, Klamt F, Vianna MM, Schröder N, Quevedo J, Benfato MS, Moreira JC, Walz R. Lipid peroxidation in hippocampus early and late after status epilepticus induced by pilocarpine or kainic acid in Wistar rats. *Neurosci Lett.* 2000 Sep 22;291(3):179-82.
16. de Lima E, Soares JM, del Carmen Sanabria Y, Gomes Valente S, Priel MR, Chada E, Baracat E, Cavalheiro E, Naffah-Mazzacoratti M, Amado D, Effects of pinealectomy and the treatment with melatonin on the temporal lobe epilepsy in rats. *Brain Res* 2005; 10: 24-31.
17. Devi PU, Manocha A., Vohora D., Seizures, antiepileptics, antioxidants and oxidative stress: an insight for researchers. *Expert Opin Pharmacother* 2008 9: 3169-3177.
18. Devinsky, O., 2004. Effects of Seizures on Autonomic and Cardiovascular Function. *Epilepsy. Curr.* 4, 43-46.
19. Galano A, Tan D X, Reiter J. R. Melatonin as a natural ally against oxidative stress: a physicochemical examination *J. Pineal Res.* 2011; 51:1-16
20. Gonzalez MF, Shiraishi K, Hisanaga K, Sagar SM, Mandabach M, Sharp FR. Heat shock proteins as markers of neural injury. *Mol Brain Res.* 1989 6:93-100.
21. Gönenç S, Uysal N, Açıkgöz O, Kayatekin BM, Sönmez A, Kiray M, Aksu I, Güleçer B, Topçu A, Semin I. Effects of melatonin on oxidative stress and spatial memory impairment induced by acute ethanol treatment in rats. *Physiol Res.* 2005;54(3):341-8.
22. Greenwood, S.R., Meeker, R., Sullivan, H., Hayward, J.N., 1989. Kindling in spontaneous hypertensive rats. *Brain Res.* 495, 58-65.
23. Guisti P, Lipartiti M, Franceschini D, Schiavo N, Floream M, Manev H, Neuroprotection by melatonin from kainate-induced excitotoxicity in rats. *FASEB J* 1996; 10: 891-896.
24. Gupta YK, Briyal S. Protective effect of vinetrol against kainic acid induced seizures, oxidative stress and on the expression of heat shock proteins in rats. *Eur Neuropsychopharmacol.* 2006 Feb;16(2):85-91.
25. Hamet P, Malo D, Tremblay J. Increased transcription of a major stress gene in spontaneously hypertensive mice. *Hypertension.* 1990 Jun;15(6 Pt 2):904-8. A
26. Hamet P, Tremblay J, Malo D, Kunes J, Hashimoto T. Genetic hypertension is characterized by the abnormal expression of a gene localized in major histocompatibility complex HSP70. *Transplant Proc.* 1990, 22:2566-7. B
27. Harris ED. Regulation of antioxidant enzymes. *FASEB J.* 1992, 6(9), 2675-83;

28. Hilz, M., Devinsky, O., Doyle, W., Mauere, A., Dütsch, M., 2002. Decrease of cardiovascular modulation after temporal lobe epilepsy surgery. *Brain* 125, 985-995.
29. Hirata H, Asanuma M, Cadet JL, Melatonin attenuates methamphetamine induced toxic effects on dopamine and serotonin terminals in mouse brain. *Synapse* 1998; 30: 150–155.
30. Kubera M, Budziszewska B, Jaworska-Feil L, Basta-Kaim A, Leśkiewicz M, Tetich M, Maes M, Kenis G, Marciniak A, Czuczwar SJ, Jagła G, Nowak W, Lasoń W. Effect of topiramate on the kainate-induced status epilepticus, lipid peroxidation and immunoreactivity of rats. *Pol J Pharmacol.* 2004 56:553-61.
31. Kukreja RC, Kontos MC, Loesser KE, Batra SK, Qian YZ, Gbur CJ Jr, Naseem SA, Jesse RL, Hess ML, Kunz WS. The role of mitochondria in epileptogenesis. *Curr Opin Neurol.* 2002; 15: 179-184.
32. Lapin IP, Mirzaev SM, Ryzon IV, Oxenkrug GF (1998), Anticonvulsant activity of melatonin against seizures induced by quinolinate, kainate, glutamate, NMDA, and pentileno-tetrazole in mice. *J Pineal Res* 24:215–218.
33. Lee J. Y. and Corry M. P. Metabolic Oxidative Stress-induced HSP70 gene expression is mediated through SAPK Pathway. Role of Bcl-2 and c-Jun NH2-terminal kinase. 1998 *The Journal of Biological Chemistry*, 273, 29857-29863.
34. Levonen A-L, Vähäkangas E, Koponen J K., Ylä-Herttua S, Antioxidant Gene Therapy for Cardiovascular Disease Current Status and Future Perspectives. *Circulation.* 2008; 117: 2142-2150.
35. Liang LP, Patel M. Seizure-induced changes in mitochondrial redox status. *Free Radic Biol Med.* 2006 Jan 15;40(2):316-22. Epub 2005 Oct 14.
36. López-Meraz ML, González-Trujano ME, Neri-Bazán L, Hong E, Rocha LL. 5-HT1A receptor agonists modify epileptic seizures in three experimental models in rats. *Neuropharmacology.* 2005 Sep;49(3):367-75.
37. Lukashev ME, Klimanskaya IV, Postnov YV. Synthesis of heat-shock proteins in cultured fibroblasts from normotensive and spontaneously hypertensive rat embryos. *J Hypertens Suppl.* 1991 9(6):S182-3.
38. Mevissen M, Ebert U, Anticonvulsant effects of melatonin in amygdala-kindled rats. *Neurosci Lett* 1998; 257:13–16.
39. Milatovic D, Zivin M, Gupta RC, Dettbarn WD. Alterations in cytochrome c oxidase activity and energy metabolites in response to kainic acid-induced status epilepticus. *Brain Res.* 2001 Aug 31;912(1):67-78.
40. Mohanan PV, Yamamoto HA. Preventive effect of melatonin against brain mitochondria DNA damage, lipid peroxidation and seizures induced by kainic acid. *Toxicol Lett.* 2002 129:99-105.
41. Molina-Carballo A, Muñoz-Hoyos A, Reiter RJ, Sánchez-Forte M, Moreno-Madrid F, Rufo-Campos M, Molina-Font JA, Acuña-Castroviejo D, Utility of high doses of melatonin as adjunctive anticonvulsant therapy in a child with severe myoclonic epilepsy: two years' experience. *J Pineal Res* 1997; 23: 97-105.
42. Morales-García JA, Luna-Medina R, Martínez A, Santos A, Pérez-Castillo A. Anticonvulsant and neuroprotective effects of the novel calcium antagonist NP04634 on kainic acid-induced seizures in rats. *J Neurosci Res.* 2009 Dec;87(16):3687-96. doi: 10.1002/jnr.22165.
43. Musshoff U, Speckmann EJ. Diurnal actions of melatonin on epileptic activity in hippocampal slices of rats. *Life Sci.* 2003 Oct 3;73(20):2603-10.
44. Polizio AH, Peña C. Effects of angiotensin II type 1 receptor blockade on the oxidative stress in spontaneously hypertensive rat tissues. *Regul Pept.* 2005 May 15;128(1):1-5.
45. Rajdev S, Sharp FR. Stress proteins as molecular markers of neurotoxicity. *Toxicol Pathol.* 2000; 28:105-12.
46. Rufo-Campos M, Melatonin and epilepsy. *Rev Neurol.* 2002; Suppl. 1: S51-S58.

47. Russel J. Reiter Dun-xian Tan Carmen Osuna Eloisa Gitto Actions of Melatonin in the Reduction of Oxidative Stress *J Biomed Sci* 2000;7:444–458
48. Sabbatini, M., Catalani, A., Consoli, C., Marletta, N., Tomassoni, D., Avola, R., 2002. The hippocampus in spontaneously hypertensive rats: an animal model of vascular dementia? *Mechan. Ageing. Dev.* 123, 547–559.
49. Scorza, F.A, Arida, R.M., Cysneiros, R.M., Scorza, C.A., de Albuquerque, M., Cavaleiro, E.A., 2005. Qualitative study of hippocampal formation in hypertensive rats with epilepsy. *Arq. Neuropsiquiatr* 63, 283-288.
50. Schiffrin EL. Antioxidants in Hypertension and Cardiovascular Disease *Molecular Interventions* 2010 10 354-362
51. Shimosaka S, So YT, Simon RP. Distribution of HSP72 induction and neuronal death following limbic seizures. *Neurosci Lett.* 1992; 138: 202-206.
52. Shin EJ, Jeong JH, Chung YH, Kim WK, Ko KH, Bach JH, Hong JS, Yoneda Y, Kim HC. Role of oxidative stress in epileptic seizures. *Neurochem Int.* 2011 Aug;59(2):122-37
53. Schwob JE, Fuller T, Price JL, Olney JW. Widespread patterns of neuronal damage following systemic or intracerebral injections of kainic acid: a histological study. *Neuroscience.* 1980;5(6):991-1014.
54. Sperk G, Lassmann H, Baran H, Kish SJ, Seitelberger F, Hornykiewicz O. Kainic acid induced seizures: neurochemical and histopathological changes. *Neuroscience.* 1983 10(4):1301-15.
55. Srivastava N., Seth, K. Srivastava N, Khanna V.K., Agrawal A.K., Functional restoration using basic fibroblast growth factor (bFGF) infusion in kainic acid induced cognitive dysfunction in rat: neurobehavioural and neurochemical studies, *Neurochem. Res.* 33 (2008) 1169–1177.
56. Tan DX, Manchester LC, Sanchez-Barcelo E, Mediavilla MD, Reiter RJ. Significance of high levels of endogenous melatonin in Mammalian cerebrospinal fluid and in the central nervous system. *Curr Neuropharmacol.* 2010 Sep;8(3):162-7.
57. Tchekalarova, J., Pechlivanova, D., Itzev, D., Lazarov, N., Markova, P., Stoynev, A., 2010. Diurnal rhythms of spontaneous recurrent seizures and behavioural alterations of Wistar and spontaneously hypertensive rats in kainate model of epilepsy. *Epilepsy Behav.* 17, 23-32.
58. Tchekalarova, J., Pechlivanova, D., Atanasova Ts., Markova, P., Lozanov, V., Stoynev, A., 2011. Diurnal variations of depressive-like behavior of Wistar and spontaneously hypertensive rats in kainate model of temporal lobe epilepsy. *Epilepsy Behav.* 20, 277-285.
59. Tchekalarova, J., Petkova, Z., Pechlivanova, D., Moyanova, S., Kortenska, L., Mitreva, R., Lozanov, V., Atanasova, D., Lazarov, N., Stoynev, Al., 2013 Prophylactic treatment with melatonin after status epilepticus: Effects on epileptogenesis, neuronal damage and behavioral changes in kainate model of temporal lobe epilepsy. *Epilepsy Behav.* 27, 174-187.
60. Tomson, T., Ericson, M., Ihrman, C., Lindblad, L.E., 1998. Heart rate variability in patients with epilepsy. *Epilepsy Res.* 30, 77–83.
61. Vass K, Berger ML, Nowak TS Jr, Welch WJ, Lassmann H. Induction of stress protein HSP70 in nerve cells after status epilepticus in the rat. *Neurosci Lett.* 1989 100(1-3):259-64.
62. Vogt M., Bauer MKA., Ferrari D., Schulze-Osthoff K. Oxidative stress and hypoxia/reoxygenation trigger CD95 (APO-1/Fas) ligand expression in microglial cells
63. *FEBS Lett.* 1998, 429(1), 67-72.
64. Vorobyov, V., Schibaev, N., Kaptsov, V., Kovalev, G., Sengpiel, F., 2011. Cortical and hippocampal EEG effects of neurotransmitter agonists in spontaneously hypertensive vs. kainate-treated rats. *Brain Res.*1383, 154-68.
65. Waldbaum S, Patel M. Mitochondrial dysfunction and oxidative stress: a contributing link to acquiredepilepsy? *J Bioenerg Biomembr.* 2010 Dec;42(6):449-55
66. Wills E. D. Lipid peroxide formation in microsomes. General considerations *Biochem J.* 1969 June; 113(2): 315–324.

67. Xu K, Stringer JL. Antioxidants and free radical scavengers do not consistently delay seizure onset in animal models of acute seizures. *Epilepsy Behav.* 2008 Jul;13(1):77-82.

68. Yamamoto HA, Tang HW, Melatonin attenuates l-cysteine-induced seizures and peroxidation lipid in the brain of mice. *J Pineal Res* 1996; 21:108–113.

69. Yonei Y, Hattori A, Tsutsui K, Okawa M, Ishizuka B. Effects of Melatonin: Basics Studies and Clinical Applications. *Anti-Aging Medicine* 7 (7) : 85-91, 2010

70. Yoshikazu Yonei, Atsuhiko Hattori, Kazuyoshi Tsutsui, Masako Okawa, Bunpei Ishizuka Effects of Melatonin: Basics Studies and Clinical Applications *Anti-Aging Medicine* 7 (7) : 85-91, 2010

71. Zhou J, Ando H, Macova M, Dou J, Saavedra JM. Angiotensin II AT1 receptor blockade abolishes brain microvascular inflammation and heat shock protein responses in hypertensive rats. *J Cereb Blood Flow Metab.* 2005; 25(7):878-86.

Table 1

Effect of pretreatment with melatonin infused subcutaneously via osmotic minipumps for 14 days (10 mg/kg per day) on KA- induced seizures in Wistar and spontaneously hypertensive rats (SHRs). Analysis of data by two-way ANOVA indicated a main Strain effect [$F_{1,34} = 10.014$, $p < 0.003$] and Strain x Drug interaction [$F_{1,34} = 5.129$, $p < 0.031$] for the latency of KA-induced seizures. $^{\circ}P = 0.02$ versus Wistar-KA-veh group; $**P = 0.034$ versus SHRs-KA-veh group (Kruskal-Wallis test)

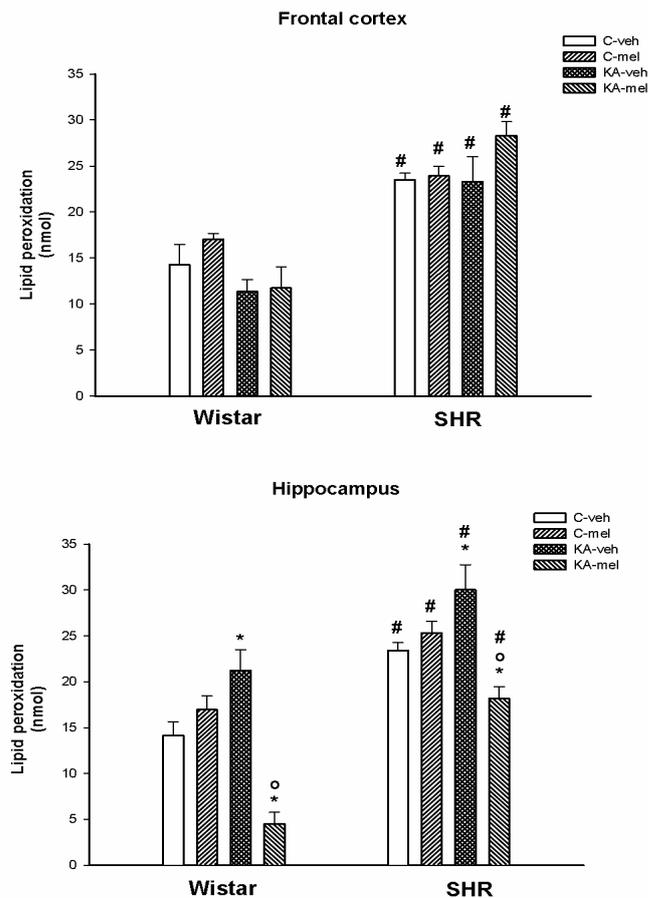


Fig. 1

Lipid peroxidation in the frontal cortex (A) and the hippocampus (B) of controls (C) or KA-treated Wistar and SHRs infused with vehicle (veh) or melatonin (mel) (details in the text to Table 1). Data are presented as means \pm SEM (n = 10). Analysis of data by three-way ANOVA indicated a main Strain effect [$F_{1,63}= 69.337$, $p<0.001$] in (A), a main Strain effect [$F_{1,63}= 66.905$, $p<0.001$], a main Drug effect [$F_{1,63}=23.638$, $p<0.001$] and KA-treatment x Drug interaction [$F_{1,63}= 46.070$, $p<0.001$] in (B). * $p < 0.05$ vs C-veh group; $^{\circ}p < 0.05$ vs KA-veh group, $^{\#}p < 0.05$ vs Wistar rats.

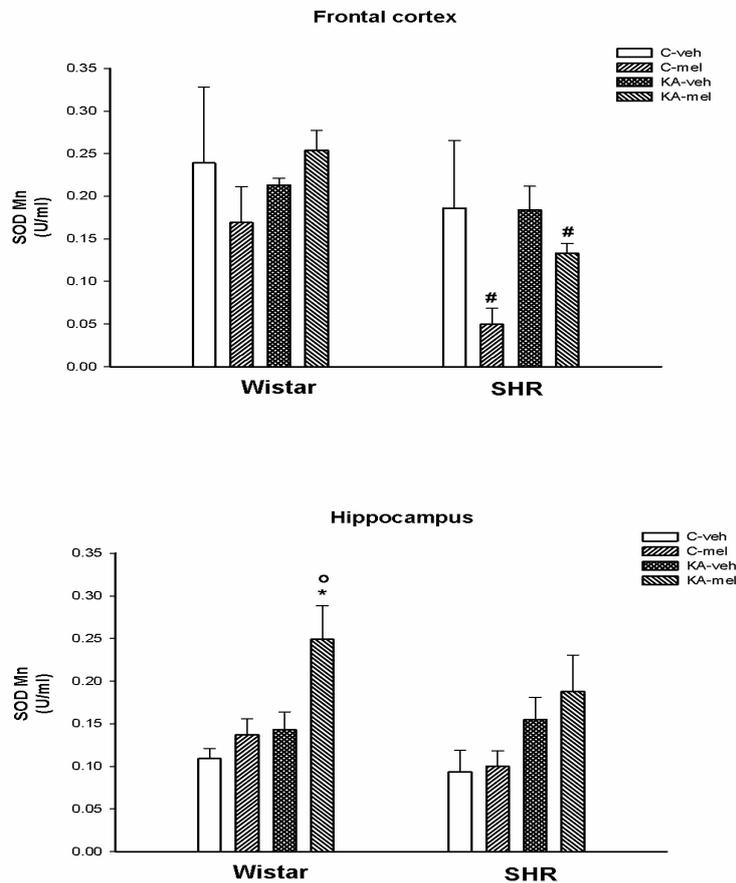


Fig. 2

Cytosolic superoxide dismutase (SOD Cu/Zn) activity in the frontal cortex (A) and the hippocampus (B) of controls (C) or KA-treated Wistar and SHRs pretreated with either vehicle (veh) or melatonin (mel) (details in Table 1). Data are presented as means \pm SEM (n = 10). Analysis of data by three-way ANOVA indicated a main Strain effect [$F_{1,61}= 49.083$, $p<0.001$], a main KA-treatment effect [$F_{1,61}= 42.620$, $p<0.001$] and Strain x KA-treatment x Drug interaction [$F_{1,61}= 42.139$, $p<0.001$] in (A). Analysis by three-way ANOVA showed a main KA-treatment effect [$F_{1,62}= 19.777$, $p<0.001$], a main Drug effect [$F_{1,62}= 27.248$, $p<0.001$], interaction Strain x KA-treatment x Drug [$F_{1,62}= 9.357$, $p=0.003$] in (B). * $p < 0.05$ vs C-veh group; $^{\circ}p < 0.05$ vs KA-veh group, $^{\#}p < 0.05$ vs Wistar rats.

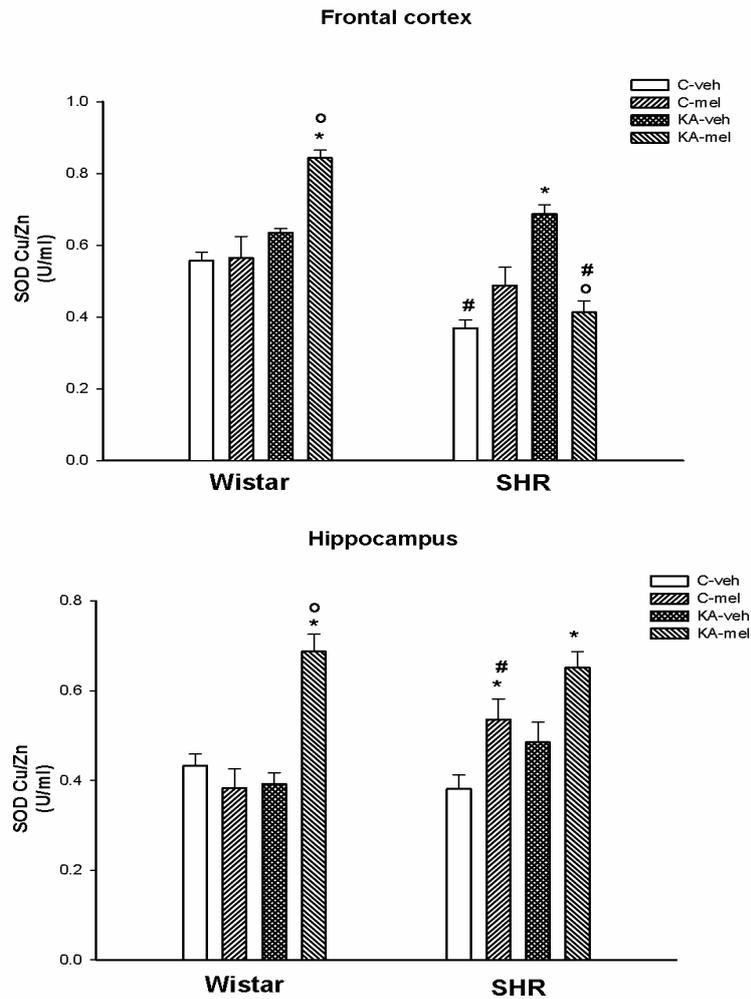
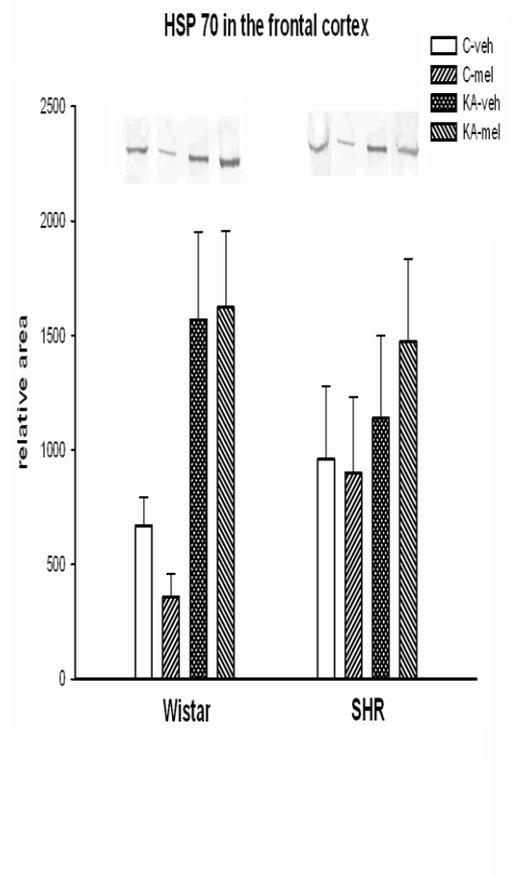


Fig. 3

Mitochondrial superoxide dismutase (SOD Mn) activity in the frontal cortex (A) and the hippocampus (B) of controls (C) or KA-treated Wistar and SHRs pretreated with either vehicle (veh) or melatonin (mel) (details in Table 1). Data are means \pm SEM (n = 10). Analysis of data by three-way ANOVA indicated a main KA-treatment effect [$F_{1,68} = 14.472$, $p < 0.001$] in (B) and main Strain effect [$F_{1,62} = 7.979$, $p = 0.007$] in (A). * $p < 0.05$ vs C-veh group; ° $p < 0.05$ vs KA-veh group, # $p < 0.05$ vs Wistar rats.

**Fig. 4**

Heat shock protein (HSP) 70 in the frontal cortex (A) and the hippocampus (B) of controls (C) or KA-treated Wistar and SHRs pretreated with either vehicle (veh) or melatonin (mel) (details in Table 1). Data are means \pm SEM (n = 10). Analysis of data by three-way ANOVA indicated a main KA-treatment effect [$F_{1,55} = 9.308$, $p = 0.004$] in (A), a main Strain effect [$F_{1,52} = 18.725$, $p < 0.001$], a main KA-treatment effect [$F_{1,52} = 18.167$, $p < 0.001$] and interaction Strain x KA-treatment x Drug [$F_{1,52} = 5.088$, $p = 0.029$]. * $p < 0.05$ vs C-veh group; $^{\circ}p < 0.05$ vs KA-veh group, $^{\#}p < 0.05$ vs Wistar rats.